











### Anti-VEGF Timeline: From Conception to Delivery

<b>1913</b>	 A. Carrel et al. hypothesized the existence of growth factors for cellular tissues. <sup>i</sup>
<b>1939</b>	 A.G. Ide et al. observed that transplanted tumors induced neovascular growth and postulated a tumor-derived “blood vessel growth stimulating factor”. <sup>ii</sup>
<b>1945</b>	 G.H. Algire et al. suggested that tumor growth depends on a rich vascular supply. <sup>iii</sup>
<b>1948</b>	 I.C. Michaelson suspected that a diffusible angiogenic factor “factor X” existed in the retina. <sup>iv</sup>
<b>1971</b>	 J. Folkman published an influential paper on the implications of anti-angiogenesis. <sup>v</sup>
<b>1983</b>	 D.R. Senger et al. identified and characterized vascular permeability factor (VPF). <sup>vi</sup>
<b>1989</b>	 N. Ferrara et al. cloned, sequenced and characterized VEGF (previously known as VPF). <sup>vii,viii</sup>
<b>1992</b>	 Two studies showed that VEGF mRNA expression was induced by hypoxia. <sup>ix,x</sup>
<b>1993</b>	 K.J. Kim et al. identified monoclonal antibodies that can target VEGF and inhibit tumor growth. <sup>xi</sup>
<b>1994</b>	 J.W. Miller et al. demonstrated in primates that hypoxic retina can produce VEGF and that VEGF is temporally and spatially associated with iris neovascularization. <sup>xii</sup>

<b>1994</b>	<ul style="list-style-type: none"> <li>✚ L.P. Aiello et al. reported that patients with active neovascular ocular disease showed increased levels of VEGF in their ocular fluids.<sup>xiii</sup> A paper in the American Journal of Ophthalmology (AJO) by A.P. Adamis et al. showed the same thing (smaller number of patients).<sup>xiv</sup></li> </ul>
<b>1996</b>	<ul style="list-style-type: none"> <li>✚ Two articles reported the presence of VEGF in choroidal neovascular membranes from patients with wet AMD.<sup>xv,xvi</sup></li> <li>✚ Shima et al. cloned an mRNA expression of VEGF in ischemic monkey retina.</li> <li>✚ Tolentino et al. found VEGF injection was sufficient to produce iris neovascularization and neovascular glaucoma in normal monkey eyes.<sup>xvii</sup></li> <li>✚ A.P. Adamis et al. found that anti-VEGF antibodies (precursor of Avastin) suppress iris neovascularization in a primate model.<sup>xviii</sup></li> </ul>
<b>1997</b>	<ul style="list-style-type: none"> <li>✚ Pournaras et al. showed systemic hyperoxia decreases VEGF gene expression in ischemic monkey retina.<sup>xix</sup></li> <li>✚ Phase 1 clinical trial of bevacizumab initiated by Genentech for cancer.</li> </ul>
<b>1999</b>	<ul style="list-style-type: none"> <li>✚ Kim et al. showed constitutive expression of VEGF and VEGF R1 and R2 in normal monkey eyes.<sup>xx</sup></li> <li>✚ Y. Chen et al. at Genentech developed a better diffusing variant, ranibizumab, for use in the eye.<sup>xxi</sup></li> </ul>
<b>2000</b>	<ul style="list-style-type: none"> <li>✚ Genentech initiated the first clinical trial with wet AMD subjects.</li> </ul>
<b>2002</b>	<ul style="list-style-type: none"> <li>✚ Krzystolik et al. showed prevention of experimental CNV with intravitreal anti-VEGF antibody fragment (precursor of ranibizumab).<sup>xxii</sup></li> </ul>
<b>2004</b>	<ul style="list-style-type: none"> <li>✚ Pegaptanib received FDA approval for nvAMD.</li> </ul>
<b>2006</b>	<ul style="list-style-type: none"> <li>✚ Two-year results from the MARINA trial of Ranibizumab for neovascular AMD were published by Rosenfeld et al.<sup>xxiii</sup></li> <li>✚ Ranibizumab received FDA approval for neovascular AMD.</li> </ul>

	<ul style="list-style-type: none"> <li>✚ Rosenfeld et al. published OCT evidence of the efficacy of bevacizumab in patients with nvAMD.<sup>xxiv</sup></li> </ul>
<b>2010</b>	<ul style="list-style-type: none"> <li>✚ Ranibizumab received FDA approval for retinal vein occlusion.</li> </ul>
<b>2011</b>	<ul style="list-style-type: none"> <li>✚ Aflibercept received FDA approval for neovascular AMD.</li> </ul>
<b>2012</b>	<ul style="list-style-type: none"> <li>✚ Ranibizumab received FDA approval for diabetic macular edema.</li> </ul>
<b>2014</b>	<ul style="list-style-type: none"> <li>✚ Studies on the effectiveness of ranibizumab and pegaptanib on patients with macular edema from central retinal vein occlusion commenced.</li> </ul>
<b>2015</b>	<ul style="list-style-type: none"> <li>✚ Ranibizumab &amp; aflibercept received FDA approval for treatment of DR in DME patients.</li> </ul>
<b>2019</b>	<ul style="list-style-type: none"> <li>✚ Brolucizumab-dblb received FDA approval for nAMD and DME.</li> </ul>
<b>2021</b>	<ul style="list-style-type: none"> <li>✚ A Lucentis biosimilar was the first to receive FDA approval for nAMD, macular edema following RVO and myopic CNV.</li> </ul>
<b>2022</b>	<ul style="list-style-type: none"> <li>✚ Another biosimilar received FDA approval for nAMD, DR, DME, myopic DNV and macular edema following RVO.</li> <li>✚ Faricimab-svoa received FDA approval for nAMD and DME.</li> </ul>
<b>2023</b>	<ul style="list-style-type: none"> <li>✚ Higher dose Aflibercept received FDA approval for wet AMD, DME and DR.</li> <li>✚ Aflibercept received FDA approval for treatment for preterm infants with retinopathy of prematurity.</li> <li>✚ Faricimab-svoa received FDA approval for the treatment of RVO.</li> </ul>

## References

- <sup>i</sup> Carrel A. Artificial activation of the growth in vitro of connective tissue. *J Exp Med.* 1913; 17: 14–19.
- <sup>ii</sup> Ide A.G., Baker N.H., Warren S.L. Vascularization of the Brown Pearce rabbit epithelioma transplant as seen in the transparent ear chamber. *Am J Roentgenol.* 1939; 42: 891–899.
- <sup>iii</sup> Algire G.H., Chalkley H.W., Legallais F.Y., Park H.D. Vascular reactions of normal and malignant tissues in vivo. I. Vascular reactions of mice to wounds and to normal and neoplastic transplants. *J Natl Cancer Inst.* 1945; 6: 73–85.
- <sup>iv</sup> Michaelson I.C. The mode of development of the vascular system of the retina with some observations on its significance for certain retinal diseases. *Trans Ophthalmol Soc U K.* 1948; 68: 137–180.
- <sup>v</sup> Folkman J. Tumor angiogenesis: therapeutic implications. *N Engl J Med.* 1971; 285: 1182–1186.
- <sup>vi</sup> Senger D.R., Galli S.J., Dvorak A.M., Perruzzi C.A., Harvey V.S., Dvorak H.F. Tumor cells secrete a vascular permeability factor that promotes accumulation of ascites fluid. *Science.* 1983; 219: 983–985.
- <sup>vii</sup> Ferrara N., Henzel W.J. Pituitary follicular cells secrete a novel heparin-binding growth factor specific for vascular endothelial cells. *Biochem Biophys Res Commun.* 1989; 161: 851–858.
- <sup>viii</sup> Leung D.W., Cachianes G., Kuang W.J., Goeddel D.V., Ferrara N. Vascular endothelial growth factor is a secreted angiogenic mitogen. *Science.* 1989; 246: 1306–1309.
- <sup>ix</sup> Shweiki D., Itin A., Soffer D., Keshet E. Vascular endothelial growth factor induced by hypoxia may mediate hypoxia-initiated angiogenesis. *Nature.* 1992; 359: 843–845.
- <sup>x</sup> Plate K.H., Breier G., Weich H.A., Risau W. Vascular endothelial growth factor is a potential tumour angiogenesis factor in human gliomas in vivo. *Nature.* 1992; 359: 845–848.
- <sup>xi</sup> Kim K.J., Li B., Winer J., et al. Inhibition of vascular endothelial growth factor-induced angiogenesis suppresses tumor growth in vivo. *Nature.* 1993; 362: 841–844.
- <sup>xii</sup> Miller J.W., Adamis A.P., Shima D.T., D'Amore P.A., Moulton R.S., O'Reilly M.S., Folkman J., Dvorak H.F., Brown L.F., Berse B., Yeo T.K., Yeo K.T. Vascular endothelial growth factor/vascular permeability factor is temporally and spatially correlated with ocular angiogenesis in a primate model. *Am J Pathol.* 1994;145:574–584.
- <sup>xiii</sup> Aiello L.P., Avery R.L., Arrigg P.G., Keyt B.A., Jampel H.D., Shah S.T., Pasquale L.R., Thieme H., Iwamoto M.A., Park J.E., Nguyen H.V., Aiello L.M., Ferrara N., King G.L. Vascular endothelial growth factor in ocular fluid of patients with diabetic retinopathy and other retinal disorders. *N Engl J Med.* 1994;331:1480–1487.
- <sup>xiv</sup> Adamis A.P., Miller J.W., Bernal M.T., D'Amico D.J., Folkman J., Yeo T.K., Yeo K.T. Increased vascular endothelial growth factor levels in the vitreous of eyes with proliferative diabetic retinopathy. *Am J Ophthalmol.* 1994 Oct 15;118(4):445-50.
- <sup>xv</sup> Lopez P.F., Sippy B.D., Lambert H.M., Thach A.B., Hinton D.R. Transdifferentiated retinal pigment epithelial cells are immunoreactive for vascular endothelial growth factor in surgically excised age-related macular degeneration-related choroidal neovascular membranes. *Invest Ophthalmol Vis Sci.* 1996; 37: 855–868.
- <sup>xvi</sup> Kvant A., Algere P.V., Berglin L., Seregard S. Subfoveal fibrovascular membranes in age-related macular degeneration express vascular endothelial growth factor. *Invest Ophthalmol Vis Sci.* 1996; 37: 1929–1934.
- <sup>xvii</sup> Tolentino M.J., Miller J.W., Gragoudas E.S., Chatzistefanou K., Ferrara N., Adamis A.P. Vascular endothelial growth factor is sufficient to produce iris neovascularization and neovascular glaucoma in a nonhuman primate. *Arch Ophthalmol.* 1996 Aug;114(8):964-70.
- <sup>xviii</sup> Adamis A.P., Shima D.T., Tolentino M.J., Gragoudas E.S., Ferrara N., Folkman J., D'Amore P.A., Miller J.W. Inhibition of vascular endothelial growth factor prevents retinal ischemia-associated iris neovascularization in a nonhuman primate. *Arch Ophthalmol.* 1996;114:66–71.
- <sup>xix</sup> Pournaras C.J., Miller J.W., Gragoudas E.S., Husain D., Munoz J.L., Tolentino M.J., Kuroki M., Adamis A.P. Systemic hyperoxia decreases vascular endothelial growth factor gene expression in ischemic primate retina. *Arch Ophthalmol.* 1997 Dec;115(12):1553-8.

---

<sup>xx</sup> Kim I., Ryan A.M., Rohan R., Amano S., Agular S., Miller J.W., Adamis A.P. Constitutive Expression of VEGF, VEGFR-1, and VEGFR-2 in Normal Eyes. *Investigative Ophthalmology & Visual Science* August 1999, Vol.40, 2115-2121.

<sup>xxi</sup> Chen Y., Wiesmann C., Fuh G., Li B., Christinger H.W., McKay P., de Vos A.M., Lowman H.B. Selection and analysis of an optimized anti-VEGF antibody: crystal structure of an affinity-matured Fab in complex with antigen. *J Mol Biol.* 1999;293:865–881.

<sup>xxii</sup> Krzystolik M.G., Afshari M.A., Adamis A.P., Gaudreault J., Gragoudas E.S., Michaud N.A., Li W., Connolly E., O'Neill C.A., Miller J.W. Prevention of experimental choroidal neovascularization with intravitreal anti-vascular endothelial growth factor antibody fragment. *Arch Ophthalmol.* 2002 Mar;120(3):338-46.

<sup>xxiii</sup> Rosenfeld P.J., Brown D.M., Heier J.S., et al. Ranibizumab for neovascular age-related macular degeneration. *N Engl J Med.* 2006; 355: 1419–1431.

<sup>xxiv</sup> Rosenfeld P.J., Moshfeghi A.A., Puliafito C.A. Optical coherence tomography findings after an intravitreal injection of bevacizumab (Avastin) for neovascular age-related macular degeneration. *Ophthalmic Surg Lasers Imaging.* 2005;36:331–335.